



Tryptophan Metabolism in Neurodevelopment and Its Implications For Neurodevelopmental Disorders

Maria Grazia Giuliano¹ · Paola Tognini¹

Received: 19 April 2025 / Accepted: 22 August 2025
© The Author(s) 2025

Abstract

The role of tryptophan metabolism has been recognized in a wide range of physiological and pathological processes but is still only partially understood. Growing evidence highlights the importance of maintaining tryptophan homeostasis throughout life, with its disruption now linked to various neuropsychiatric conditions spanning from early life to aging. While it is increasingly evident that alterations in tryptophan metabolism have significant implications for both neurodevelopmental and neurodegenerative disorders, research has predominantly focused on the latter, leaving neurodevelopmental aspects comparatively underexplored. This review provides a comprehensive overview of both preclinical and clinical studies, highlighting the intricate relationship between tryptophan metabolism and neurodevelopment. Particular focus is given to the kynurenine pathway and gut microbiota-derived indole production, two interconnected metabolic branches with profound effects on brain maturation, plasticity, and immune regulation. Finally, we examine the pathophysiological consequences of tryptophan dysregulation in neurodevelopmental disorders, including autism spectrum disorder, attention-deficit/hyperactivity disorder, and Rett syndrome. We also discuss potential therapeutic strategies targeting tryptophan metabolism in these conditions.

Keywords Tryptophan · Kynurenine · Indole · Gut microbiota · Neurodevelopment · Neurodevelopmental disorders

Introduction: Tryptophan Metabolism as a Key Source of Neuroactive Compounds Regulated by Endogenous and Exogenous Factors

L-Tryptophan (Trp), the least abundant amino acid in the human body, contains an indole ring that animals cannot synthesize, which must be obtained through diet or protein degradation [1]. Once released from dietary proteins, Trp is converted into indole by several phyla of gut bacteria, producing neuroactive indole derivatives that influence brain function through a partially unknown mechanism [2–5]. The remaining Trp serves as a precursor of serotonin (5-hydroxytryptamine, 5-HT), a key neurotransmitter/neuromodulator within the enteric and central nervous systems (CNS), primarily produced in gut enterochromaffin cells

[6, 7]. Alternatively, Trp and indoles are absorbed through the intestinal epithelium and delivered to the hepatic portal system [8]. In the liver, indole is oxidized and sulphated to indoxyl sulfate, a neurotoxic compound that, if in excess, can cause the accumulation of neurotransmitters which block the efflux transporter in the blood-brain barrier (BBB) [9]. Only 5% of Trp that reaches the liver is used for protein synthesis, while the other 95% follows the kynurenine pathway (KP), producing a range of neuroactive intermediates collectively known as kynurenines (Fig. 1).

These intermediates have been shown to influence brain function, with imbalances linked to various neurological disorders such as schizophrenia, depression, Alzheimer's disease, and epilepsy [4, 10–16]. Unmetabolized Trp enters the bloodstream, either albumin-bound or free, with the latter crossing the BBB via L-type amino acid transporters (LAT) [17]. In the brain, Trp undergoes metabolism for (i) kynurenines, (ii) nicotinamide adenine dinucleotide (NAD⁺), (iii) 5-HT (10–20%), and (iv) melatonin [18]. Together, these biochemical cascades influence mood regulation, stress responses, sleep, immune function, and overall neurological health [19–21] (Fig. 2). Trp deficiency impairs protein synthesis, causing muscle and weight loss [18]. Due to its

✉ Maria Grazia Giuliano
maria.giuliano@santannapisa.it

✉ Paola Tognini
paola.tognini@santannapisa.it

¹ Health Science Interdisciplinary Center, School of Advanced Studies, Pisa, Italy

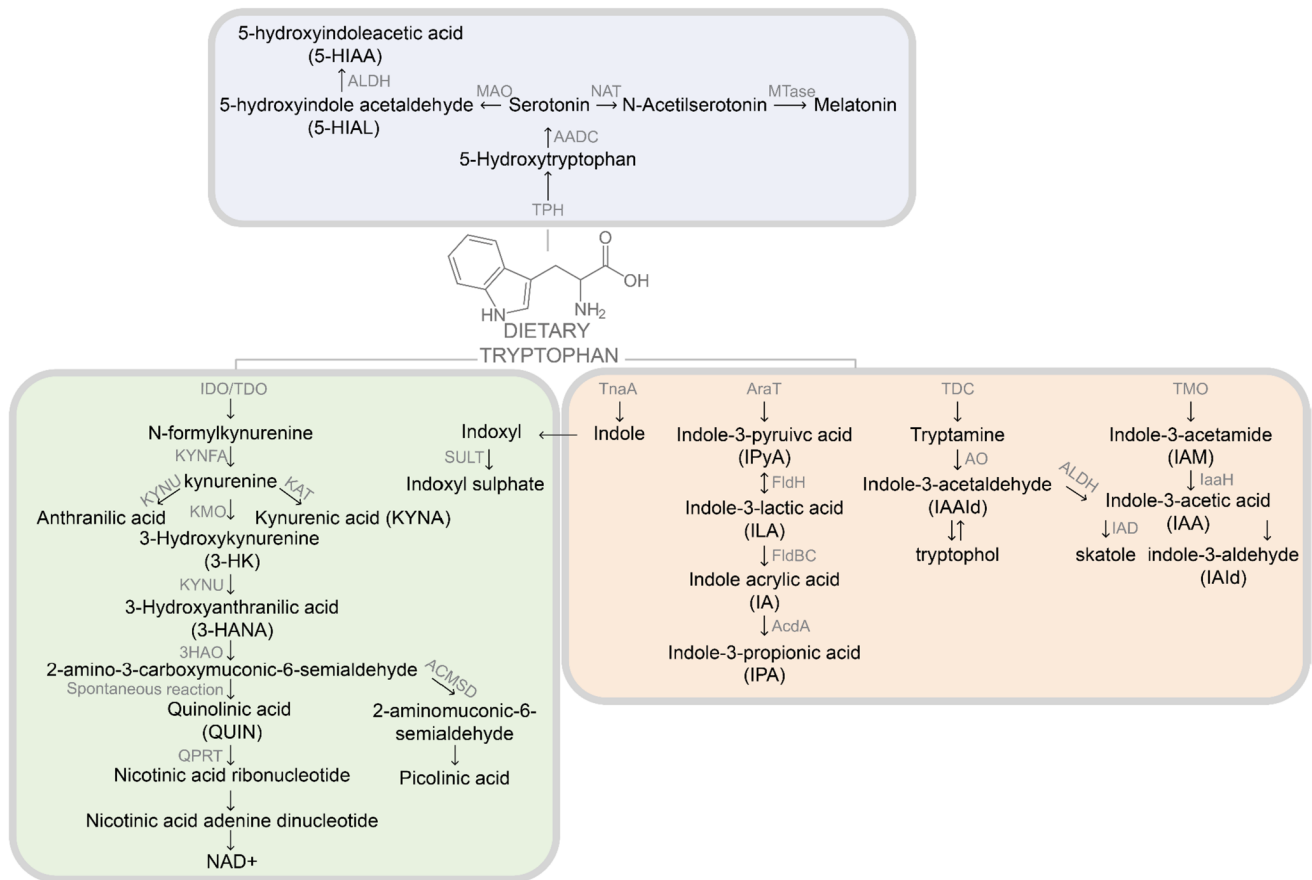


Fig. 1 Overview of major tryptophan metabolic pathways. Tryptophan (Trp) is metabolized via three major routes: the serotonin/melatonin pathway (top, blue box), the kynurenine pathway (left, green box), and the microbial indole pathway (right, orange box). Key enzymes are indicated along each pathway. TPH, tryptophan hydroxylase; AADC, aromatic L-amino acid decarboxylase; NAT, aralkylamine N-acetyltransferase; MTase, methyltransferase; MAO, monoamine oxidase; ALDH, aldehyde dehydrogenase; IDO, indoleamine 2,3-dioxygenase; TDO, tryptophan 2,3-dioxygenase; KMO, kynurenine 3-monooxygenase; KYNU, kynureninase; KAT, kynurenine aminotransferase; QPRT, quinolinate phosphoribosyltransferase; ACMSD, aminocarboxymuconate semialdehyde decar-

boxylase; TnaA, tryptophanase; SULF, sulfotransferase; AraT, aromatic amino acid transaminase; FldH/FldBC, aldehyde and dehydrogenase enzymes; AcdA, acyl-CoA dehydrogenase; TDC, tryptophan decarboxylase; AO, amine oxidase; TMO, tryptophan monooxygenase; IAD, indole-3-acetate decarboxylase; IaaH, indole-3-acetamide hydrolase; 5-HT, serotonin; 5-HIAA, 5-hydroxyindoleacetic acid; 5-HIAL, 5-hydroxyindole acetaldehyde; IPA, indole-3-propionic acid; IAA, indole-3-acetic acid; ILA, indole-3-lactic acid; IAAld, indole-3-acetaldehyde; IPyA, indole-3-pyruvic acid; IAld, indole-3-aldehyde; QUIN, quinolinic acid; KYNA, kynurenic acid; NAD⁺, nicotinamide adenine dinucleotide

neuroactive properties, imbalanced intake also affects the CNS, influencing mood, fatigue, and perception [22, 23]. In the Western world, excessive Trp consumption (70–200 mg/kg) is common, often through diet or supplementation with antidepressants that augment 5-HT function [24]. Such high levels of Trp intake are associated with a variety of side effects, from mild symptoms like tremor, nausea, and dizziness, to rare cases of serotonin syndrome—a severe condition marked by delirium, myoclonus, hyperthermia, and coma, requiring urgent medical care [25]. A high-fat diet is also known to negatively affect Trp metabolism through multiple mechanisms. These include immune activation, which leads to elevated cytokine levels and the accumulation of neurotoxic kynurenine metabolites [26]; increased

activity of indoleamine 2,3-dioxygenase (IDO), the first and rate-limiting enzyme of the KP, which shifts Trp metabolism toward the kynurenine branch and may trigger inflammatory responses [27]; and modulation of the gut microbiota, with a reduction in beneficial bacterial species and an increase in Trp degradation [28]. Several pieces of evidence also indicate that fiber intake is strongly connected to Trp metabolic fate, with a central role played by the intestinal microbiota [29–32].

Trp metabolism plays a critical but incompletely understood role in health and disease. While the 5-HT/melatonin pathway is well characterized, the KP and microbiota-derived indoles have only recently gained attention [33, 34]. Emerging evidence links imbalances in these

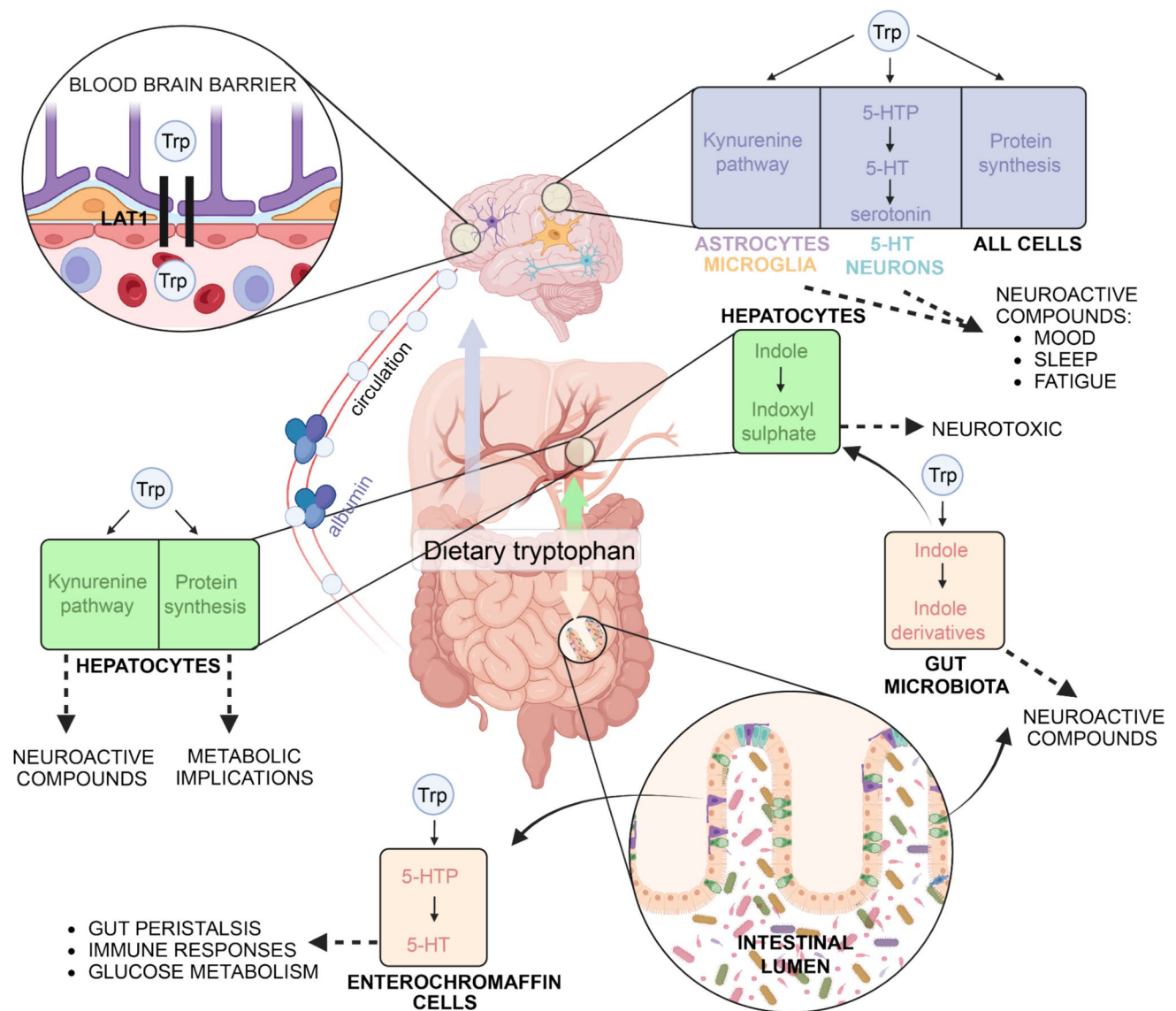


Fig. 2 Tryptophan distribution and metabolic fate across tissues. Schematic illustration of dietary tryptophan (Trp) uptake and its major metabolic pathways in different tissues, including the serotonin pathway (gut enterochromaffin cells and brain), the kynurenine pathway (liver), and microbiota-mediated indole pathway (gut microbiota, liver). Briefly, Trp is transported across the blood–brain barrier via LAT1 (L-type amino acid transporter 1) and converted into serotonin (5-HT) through the intermediate 5-hydroxytryptophan (5-HTP). In peripheral tissues, Trp is largely catabolized via the kynurenine path-

way, generating various neuroactive and immunomodulatory metabolites. In the gut, microbial enzymes convert Trp into a wide range of indole derivatives. Solid black arrows represent enzymatic steps or metabolic intermediates within each pathway. Dashed lines indicate physiological roles or downstream effects of Trp-derived metabolites. Trp, tryptophan; 5-HTP, 5-hydroxytryptophan; 5-HT, serotonin (5-hydroxytryptamine); LAT1, L-type amino acid transporter 1. Illustration created with BioRender.com

pathways to pathology, highlighting their therapeutic potential. This review examines the impact of kynurenine and indole metabolites on neurodevelopment, particularly neural function and neuroinflammation, and evaluates pre-clinical and clinical findings linking Trp metabolism to CNS health. Finally, we discuss whether targeting these pathways could offer new therapeutic strategies for neurodevelopmental disorders.

Tryptophan Metabolism in the CNS

Kynurenines have relevant and unique effects in the CNS, and evidence suggests that they play an important role in brain development [35]. L-kynurenine (kynurenine) is metabolized from Trp by the IDO1 and IDO2 enzymes and Trp 2,3-dioxygenase (TDO2), producing N-formylkynurenine, which rapidly converts to kynurenine. Kynurenine

enters the brain from circulation and is taken up by astrocytes and microglia. In astrocytes, kynurenine aminotransferase (KAT) catalyzes the irreversible transamination of kynurenine to kynurenic acid (KYNA) [36], a neuroactive antagonist of $\alpha 7$ nicotinic acetylcholine ($\alpha 7$ nACh) and *N*-methyl-D-aspartate (NMDA) receptors [37]. In microglial cells [38], the kynurenine 3-monooxygenase (KMO) metabolizes kynurenine to 3-hydroxykynurenine (3-HK), leading to 3-hydroxyanthranilic acid via kynureninase catalysis followed by quinolinic acid (QA), a neurotoxic NMDA receptor agonist that promotes free radical generation [37]. KYNA, in contrast, reduces excitotoxicity by antagonizing ionotropic glutamate receptors [39]. Approximately half of brain kynurenine is derived peripherally, linking gut and liver Trp metabolism to kynurenine levels in the CNS [37].

Trp metabolites also activate the Aryl hydrocarbon receptor (AhR), a transcription factor influenced by environmental, microbial, and metabolic signals, with key roles in metabolism, cell differentiation, intestinal barrier integrity, and immune function [40–44]. In the CNS, AhR activation in brain microvessels and the BBB, where its expression is elevated, disrupts vascular homeostasis, contributing to neurodegenerative diseases [45]. This disruption is further exacerbated by oxidative stress [46], activation of inflammatory pathways, induction of endothelial cell senescence, and vascular calcification, as seen in Alzheimer's-related cerebral amyloid angiopathy. Dysregulated AhR signaling also affects circadian rhythms, while gut microbiota dysbiosis may amplify brain pathology via Trp-AhR pathways [45, 47]. Moreover, kynurenine-mediated AhR activation is implicated in ischemic injury but may have neuroprotective effects in retinal ischemia/reperfusion injury [48]. During postnatal development, excessive AhR activation has been shown to disrupt olfactory interneuron migration and dendritic growth in mice [49, 50]. Also, AhR modulation appears essential for normal brain maturation in zebrafish embryos supporting its relevance during prenatal and early postnatal brain development. As many AhR-modulating metabolites derive from Trp, this pathway likely links immune signaling to neurodevelopment, with AhR activity potentially helping to explain the role of Trp in both brain maturation and disorder vulnerability.

Finally, the accumulation of kynurenine intermediates like QA in the CNS is also linked to Huntington's disease, schizophrenia, and other inflammatory conditions [51]. Furthermore, 3-hydroxy-anthranilic acid and 3-hydroxy-kynurenine can induce neuronal apoptosis or necrosis, contributing to neurodegeneration.

Dysregulated KP may impact postnatal neurodevelopment by interfering with the balance between neurotoxic (QA) and neuroprotective (KYNA) metabolites in response to inflammation. Given its role in inflammation, excitatory/inhibitory balance, and neuronal viability, KP could

be a promising therapeutic target for early-life neurological conditions.

Crucial Role of Tryptophan Metabolites in Neurodevelopment

Prenatal Stages of Brain Development

Dietary Trp is converted into neuroactive molecules that influence neuronal and glial function throughout life. Its importance during gestation is well established [52–54], as maternal circulating Trp supports fetal brain development by (i) contributing to protein synthesis and cell membrane formation [55], (ii) shaping the serotonergic system which impacts anxiety and depression [56], and (iii) regulating the kynurenine-cytokine pathway. Disruptions in the Trp/kynurenine ratio can predispose the fetus to neuropsychiatric conditions [57–60] and play a role in fetal immune development and rejection. [61, 62] Maternal Trp serves as a key substrate for placental kynurenine metabolism, with KP enzyme activity detected as early as the first trimester [63]. Placental Trp metabolism, a relevant player in the so-called placenta-brain axis [64], shifts throughout pregnancy, favoring 5-HT synthesis in early gestation and kynurenine production near term [52]. Notably, tight regulation of maternal 5-HT levels during the first trimester is essential. This is supported by a recent observational study involving 1115 women, which found that higher concentrations of 5-HT were associated with reduced embryonic and fetal growth, as well as an increased risk of small-for-gestational-age infants. Interestingly, this adverse outcome appears to be mitigated by elevated KYN levels, highlighting the importance of balanced Trp metabolism for healthy fetal development [65]. In line with these clinical findings, excess maternal Trp intake has been shown to cause fetal brain hyperserotonemia in rats, disrupting serotonergic system development and GH-IGF-1 signaling, thus impairing general growth [65]. Also, Trp metabolites possess antioxidant properties, suggesting a role in managing oxidative stress during pregnancy. Decreased fetal body weight as a consequence of excess Trp during gestation has also been observed in pregnant mice fed with a high L-Trp diet (L-Trp intake: 7.0 g/kg BW/day) [66].

Finally, since Trp metabolites exhibit antioxidant properties [67], the regulation of Trp metabolism could be used to control/attenuate oxidative stress during abnormal pregnancy.

During pregnancy, maternal Trp imbalance, influenced by stress and/or dietary factors, may disrupt serotonergic pathways and KP, affecting not only placenta metabolism and fetal growth, but also offspring behavior later in life. Disruption of these pathways is believed to contribute to the emotional and behavioral dysfunction often observed

in individuals exposed to prenatal stress [61, 62, 68, 69]. Indeed, prenatal stress leads to sex-specific behavioral changes in mice: females show depressive-like traits along with decreased hippocampal 5-HT levels and an increased 5-HT turnover rate in both the hippocampus and brainstem, while males exhibit anxiety-like behavior with elevated QA [69]. Additionally, studies in pregnant mice have shown that acute stress leads to a transient increase in fetal brain levels of the neuroprotective KYNA, while levels of the neurotoxic metabolites 3-hydroxykynurenine and QA remain unchanged [61, 62, 68]. KYN administration during early pregnancy also induces long-term behavioral changes in the offspring [70], further underscoring the critical role of Trp metabolism in neurodevelopment.

In addition, Trp metabolism may link maternal inflammation to fetal brain development, with diet and stress shaping the serotonergic–kynurenine balance and influencing long-term neurodevelopment. For instance, mid-pregnancy inflammation in a mouse model exposed to moderate doses of viral polyinosinic:polycytidylic acid (poly I:C) [71, 72] has been reported to upregulate Trp conversion to 5-HT in the placenta, leading to accumulation of placenta-derived 5-HT and blunted 5-HT axonal outgrowth specifically in the fetal forebrain [73]. Conversely, an *ex vivo* study on human placenta has recently shown that exposure to bacterial LPS and viral poly I:C impairs Trp homeostasis, resulting in decreased production of 5-HT and an imbalanced QA/KYNA ratio [74]. This is in line with previous evidence indicating that an inflammatory placental environment in humans is associated with upregulation of IDO1, indicating increased flux through the kynurenine branch, and concomitant downregulation of the 5-HT branch [75]. These conflicting findings highlight discrepancies in studies on placental inflammation and Trp metabolism in the placenta and fetus. While some stem from methodological differences, the system's complexity, involving metabolizing enzymes and protein transporters, makes it difficult to pinpoint exact mechanisms.

During neurodevelopment, Trp metabolism may mediate the complex gut-brain-immune crosstalk. The gut microbiota, shaped by maternal diet, stress, and inflammation, could influence the availability and fate of Trp, including its conversion into neuroactive and immunomodulatory metabolites [76–78]. These Trp-derived metabolites, in turn, may act as signaling molecules along the gut-brain axis. Embryonic Trp exposure in chicken embryos has been shown to reduce body weight and aggressive behavior in male offspring, while also altering gut microbiota composition and function, supporting the hypothesis that Trp excess during gestation leads to long-term metabolic and behavioral changes via microbiota-dependent mechanisms [52]. Chen et al. (2020) demonstrated that prenatal stress elevates placental Trp and 5-HT levels and causes long-term behavioral deficits in the

offspring. These effects were absent in germ-free (GF) mice, implicating Trp-metabolizing bacteria such as *Parasutterella* and *Bifidobacterium* as key modulators of these outcomes [79]. Supplementation of stressed dams with *Bifidobacterium dentium* mitigated maternal and fetal inflammation, restored levels of neuroprotective Trp metabolites (indole-3-propionic acid (IPA), KYNA), and improved offspring social behavior [80]. Furthermore, fecal microbial transfer from Western diet-fed obese dams to chow-fed GF lactating dams suppressed circulating Trp-derived AhR ligands (e.g., indole, indole-3-acetate) and impaired innate immune responses in the offspring [81]. These findings underscore the potential role of Trp metabolites, shaped by the maternal microbiome, in modulating fetal immune and neurodevelopmental trajectories.

Postnatal Stages of Brain Development

While evidence has demonstrated that fetal exposure to altered Trp metabolism affects brain development, the postnatal period is also a critical window, requiring tight regulation of Trp pathways for healthy brain maturation. Neonatal infections with a neurotropic influenza A virus can trigger the KP, leading to a consistent increase in the expression of IDO. This activation is also marked by a temporary rise in KYNA levels in the brains of infected mice. In genetically susceptible mice, this early-life KP activation can result in long-term deficits in sensorimotor gating [82]. Lead exposure during the lactation period leads to increased levels of both KYNA and 3-HK, along with elevated KMO activity, immediately following the exposure period. The elevated KYNA levels were found to persist into adulthood and correlated with cognitive impairments [83]. Similarly, maternal deprivation in rats alters the expression of IDO at different developmental stages. Specifically, IDO expression decreases in the hippocampus during infancy but increases in the prefrontal cortex in adulthood. This model of early-life stress also leads to depressive-like behaviors in adult rats [84], suggesting that the timing of KP alterations is critical. Finally, when the KYNA precursor, kynurenine, is administered to rat dams from gestation through the early postnatal period, it results in elevated brain KYNA levels in the offspring. These animals exhibit cognitive deficits in adulthood [85], suggesting that increased brain KYNA during a critical developmental window may have lasting negative consequences on cognition.

Overall, the current literature highlights how multiple factors, including maternal diet, psychological stress, gut health and inflammation—all deeply interconnected and capable of influencing each other—may impact Trp metabolism homeostasis, with direct consequences on fetal brain development and long-term repercussions later in life [86]. Despite the complexity of Trp metabolism and its many derivatives, this

opens promising therapeutic opportunities that could potentially be implemented during prenatal life.

Consequences of Disrupted Tryptophan Metabolism: Foundations for Neurodevelopmental Disorders

Trp metabolites are critical for brain development, and their dysregulation may contribute to neurodevelopmental disorders, making this a relevant area for clinical research (Table 1). The KP, regulated by the inflammation-sensitive

enzyme IDO, links Trp metabolism to immune function. Chronic low-grade inflammation is increasingly recognized as a factor in neurodevelopmental disorders [100] highlighting the importance of Trp metabolism in neuroimmune interactions.

Furthermore, preclinical and clinical evidence links the KP to epilepsy, largely due to the properties of KYNA and QA. QA administration induces seizures in rodents [101, 102], and epilepsy-prone mice show increased QA and KP enzyme expression [103], while KYNA influences excitatory/inhibitory balance through glutamate receptors and calcium channel modulation [104]. Microbiota manipulation

Table 1 Alterations in tryptophan-derived metabolites across neurodevelopmental disorders

Disorder	Altered Trp metabolite	Tissue	Preclinical evidence	Clinical evidence	Ref
ASD	↓ Serotonin (5-HT)	Urine	-	✓	[9]
	↑ Serotonin (5-HT)	Whole blood, serum, platelets, brain	✓	✓	[87–89]
	↓ 5-Hydroxyindoleacetic acid (5-HIAA)	Urine	-	✓	[9]
	↑ 5-Hydroxyindoleacetic acid (5-HIAA)	Platelet	-	✓	[88]
	↑ Xanthurenic acid (XA)	Urine	-	✓	[9]
	↑ Quinolinic acid (QUIN)	Urine	-	✓	[9]
	↓ Kynurenic acid (KYNA)	Urine	-	✓	[9]
	↑ Indolyl 3-acetic acid (IAA)	Urine	-	✓	[9]
					[90]
	↑ Indolyl lactate (ILA)	Urine	-	✓	[9]
	↑ Indole	Feces	-	✓	[90]
	↑ Indole pyruvate (IPy)	Serum	✓	-	[89]
	↑ 3-Methylindole	Feces	-	✓	[90]
	ADHD	↑ Tryptophan (Trp)	Blood	-	✓
↑ Kynurenine (KYN)		Blood, serum	✓	✓	[91–93]
↑ Anthranilic acid		Serum, brain	✓	-	[93]
↓ Kynurenic acid (human) (KYNA)		Blood	✓	✓	[91, 92]
↑ 3-Hydroxykynurenine (mouse)		Serum, brain	✓	✓	[91–94]
↓ 3-Hydroxykynurenine (human) (3-HK)					
RTT	↓ 3-Hydroxyanthranilic acid (3-HANA)	Serum	-	✓	[95]
	↑ Kynurenine (KYN)	Brain	-	✓	[96]
	↓ Serotonin (5-HT)	Brain	✓	✓	[96, 97]
	↓ 5-Hydroxyindoleacetic acid (5-HIAA)	Brain	-	✓	[96]
	↓ Kynurenine (KYN)	Plasma	-	✓	[98]
	↓ Indole propionate (IPA)	Plasma	-	✓	[98]
	↓ Indole lactate (ILA)	Plasma	-	✓	[98]
	↑ Serotonin (5-HT)	Serum	✓	-	[99]
MeCP2 duplication syndrome	↓ Serotonin (5-HT)	Urine, cecum	✓	-	[99]
	↓ Tryptamine	Plasma	✓	-	[99]
	↓ 4-Hydroxyindole	Plasma	✓	-	[99]
	↓ 3-Indole propionate (IPA)	Plasma	✓	-	[99]
	↓ Indole lactate (ILA),	Plasma	✓	-	[99]
	↓ 5-Hydroxyindoleacetic acid (5-HIAA)	Plasma	✓	-	[99]

via antibiotics or IDO1 inhibition reduces seizures in a mouse model of infantile spasms by increasing hippocampal KYNA [105]. These findings suggest that early disruptions in KP metabolism may alter neurodevelopment and heighten susceptibility to epilepsy. Patients with epileptic spasms show reduced KYNA and KYNA/kynurenine ratio in the cerebrospinal fluid [15], while those with status epilepticus exhibit a significant Trp-KP alteration, leading to QA overproduction [106]. Additionally, the KP's first and rate-limiting enzyme, IDO, found in microglia, astrocytes, neurons, and macrophages, is upregulated by inflammatory stimuli, including cytokines, LPS, amyloid peptides, HIV proteins, and interferon gamma [107].

The following section focuses on autism spectrum disorder (ASD) and attention deficit hyperactivity disorder (ADHD), two of the most prevalent and comorbid childhood neurodevelopmental disorders [108–110]. Both have been increasingly linked to immune dysfunction, gut-brain axis alterations, and metabolic abnormalities, making them relevant for exploring Trp metabolism in the context of neurodevelopment [111–113]. Although ASD and ADHD share phenotypic overlap and comorbidity, comparing their Trp metabolic profiles is challenging due to differences in study design, sample types, and metabolites analyzed [114]. Moreover, many studies do not directly link altered Trp levels to clinical features [9, 115, 116]. Still, both conditions appear to involve shifts in Trp metabolism, marked by serotonergic and KP imbalances, often linked to immune and gut microbiota changes [117].

Rett syndrome (RTT) is also discussed to offer insight into Trp-related pathways in a genetic disorder [98]. Although evidence is limited, this condition highlights the potential link between Trp metabolism and monogenic neurodevelopmental disorders, encouraging its investigation in other diverse neurodevelopmental pathologies.

Autism Spectrum Disorders

Hyperserotonemia affects approximately one-third of individuals with ASD, who show elevated platelet and urinary levels of 5-HT and its metabolite 5-HIAA, compared to neurotypical individuals and those with intellectual disabilities outside the autism spectrum [9, 87, 88]. This peripheral increase is thought to result from heightened activity of enterochromaffin cells—the primary source of systemic serotonin. Notably, enterochromaffin cell hyperactivity has been observed in inflammatory conditions such as IBS and Crohn's disease [118, 119]. Furthermore, inducing colitis in animal models similarly elevates both 5-HT levels and enterochromaffin cell counts [120, 121]. While these findings suggest a possible link between gut inflammation and hyperserotonemia in ASD, it is also plausible that both phenomena reflect a shared underlying immune alteration

associated with ASD. Peripheral hyperserotonemia may not directly reflect central serotonergic activity, although it may still contribute to systemic immune signaling relevant to ASD pathophysiology, considering the multiple ways in which peripheral immune signals can enter the brain [122–126].

Alterations in serotonergic signaling have long been known to profoundly affect behavior in animal models. In line with this, knockouts of the 5-HT_{2B} receptor in *Drosophila*—a model for studying social behavior abnormalities—induce autism-like phenotypes, including increased social spacing and repetitive grooming behaviors [127]. In the BTBR mouse model of ASD, overexpression of the 5-HT_{1A} receptor reduces stereotyped behavior in the marble-burying test and increases time spent in the center during the open field test, suggesting a potential role in modulating both repetitive behaviors and anxiety-related features associated with ASD [128].

Beyond 5-HT, urine metabolome analyses show altered KP metabolism in ASD, with a preference for xanthurenic acid and QA over neuroprotective KYNA, which may reflect immune activation. However, the specific tissue origin of these changes (e.g., brain, gut, or peripheral immune system) remains uncertain [9, 127]. A study on Egyptian children found reduced mRNA expression of enzymes involved not only in 5-HT catabolism (MAO), but also in KYNA production (AADAT) and QA production (HAAO) in blood samples from children with ASD, compared to those with learning disabilities and healthy controls. Although transcriptional changes in peripheral blood may not directly reflect enzyme expression or function in the CNS or gut, this downregulation positively correlates with ASD risk factors (i.e., parental age, iron, and vitamin D deficiency) and negatively with ASD scores [129]. A recent investigation of lymphoblastoid cell lines from 87 autistic individuals and 78 controls found that ASD patients exhibit reduced NADH production when Trp is the sole energy source. This data may suggest that Trp metabolism is altered, but cell lines are only a proxy and may not fully reflect brain-specific metabolic dynamics [130]. Such deficits may have implications for critical molecular processes during early brain development, particularly in the first month of gestation, mitochondrial homeostasis, and neuro-immune activity. Thus, an unbalanced production of Trp metabolites, including 5-HT, QA, and KYNA—which is exclusively observed in syndromic or non-syndromic autism—may contribute to abnormal neuronal network organization. This imbalance could disrupt the typical short- and long-range cortical pathways, as well as the excitatory/inhibitory ratio seen in ASD, potentially affecting even fetal cells [9, 131, 132].

The gut microbiome also plays a crucial role in ASD-related Trp metabolism. Dysbiosis in ASD patients may favor spore-forming bacteria (e.g., *Clostridium* sp.) [133],

which are hypothesized to enhance peripheral 5-HT production via increased expression of tryptophan hydroxylase 1 (TPH1) and reduced MAO activity [133, 134]. These processes are notably active in both human and mouse neonates, which also show an increased abundance of 5-HT-producing bacteria [135]. Moreover, a reduction of bile-metabolizing *Bifidobacterium* and *Blautia* species in the gut of the BTBR T+Itpr3tf/J mouse model of ASD is associated with deficient intestinal bile acid and Trp metabolism, marked GI dysfunction, and impaired social interactions. However, the BTBR model also displays high anxiety and altered stress responses, which complicate the interpretation of behavioral findings [136]. In the offspring of the maternal immune activation (MIA) mouse model, these ASD features have been shown to be ameliorated by oral treatment with the human commensal *Bacteroides fragilis* [89], which restores the increased indole pyruvate (IPy) characterizing MIA offspring to control levels. Additionally, higher levels of indole-3-acetic acid (IAA), which inhibits the mTOR pathway promoting the development of regulatory T (Treg) cells [137], and indolyl lactate [9]—both gut microbiota-derived Trp metabolites—characterize the urinary metabolome of ASD patients, while indole and 3-methylindole are increased in their feces [90].

Overall, ASD involves a significant Trp metabolism imbalance that may be influenced by the gut microbiota. Targeting intestinal microbes could help mitigate GI and behavioral symptoms.

Attention Deficit Hyperactivity Disorder

ASD and ADHD share overlapping risk genes and environmental influences, including prenatal and early postnatal factors. While ADHD has distinct features like impulsivity, both disorders exhibit traits such as executive dysfunction, hyperfocus, social challenges, sleep disturbances, and learning disabilities [138]. Similarly to ASD, Trp metabolism is a potential molecular mechanism underlying ADHD pathophysiology, as both conditions involve inflammation and immune dysregulation. Still, the effect of Trp metabolism on ADHD remains under-characterized compared to ASD [111, 116, 139, 140].

Early links between ADHD and the serotonergic system emerged from psychostimulants like Sydnocarb, which paradoxically calms ADHD models by increasing extracellular monoamines, including 5-HT [141]. It was later found that children with ADHD undergoing Sydnocarb treatment showed increased excretion of N-1-methylnicotinamide (N-MNA), along with a higher N-MNA/5-HIAA ratio, which may suggest (but does not directly demonstrate) a shift toward KP dominance over the 5-HT pathway. Given that Sydnocarb affects multiple monoamine systems,

elevated N-MNA might alternatively reflect enhanced general monoamine turnover or oxidative stress [142].

A meta-analysis by Cavaleri et al. found that untreated ADHD patients consistently have elevated blood Trp and kynurenine levels but lower KYNA compared to controls [91, 92]. Similarly, a PTCHD1 knockout ADHD/ASD mouse model shows increased KYN, anthranilic acid, and 3-HK in serum and brain, along with hyperlocomotion, impulsivity, and impaired recognition memory [93]. ADHD patients also exhibit lower serum 3-HK levels and 3-HK/KYN ratios, which may disrupt cortical maturation, decrease NAD production [92, 94], and lower 3-hydroxyanthranilic acid (3-HAA) [95]. While these findings are promising, it remains unclear whether such metabolic changes are causal mechanisms, secondary effects, or adaptive responses.

Recent evidence indicates that methylphenidate, the most widely used first-line pharmacological treatment for ADHD, along with lisdexamfetamine [143, 144], modulates Trp metabolism. In ADHD patients, methylphenidate has been shown to significantly increase plasma levels of KYNA and xanthurenic acid [145]. In a subgroup of individuals with comorbid depressive symptoms (DS+), treatment also reduced anthranilic acid levels and urinary excretion of NADH and QA, normalizing these values to levels observed in non-depressed ADHD subjects and healthy controls [145]. Daily oscillations in indole-derived metabolites also appear to differ in DS+ ADHD subtypes. For example, hyperactive-impulsive DS+ individuals show elevated morning IAA levels, (IAA) which are halved following methylphenidate treatment. The drug also reduces elevated IPA levels and restores its normal diurnal variation [146]. Studies have reported that methylphenidate treatment alters gut microbiota composition, potentially linking changes in indole-related metabolites to microbial diversity in ADHD patients [147, 148].

These findings suggest that Trp metabolism may indicate treatment effects, offering potential biomarkers. Understanding how ADHD therapies affect all three Trp pathways (Fig. 1) may reveal new targets and aid in evaluating treatment response.

Despite these findings, research on Trp metabolism's role in ADHD remains limited, highlighting the need for further studies.

Rett Syndrome

Rett syndrome (RTT) is a rare X-linked genetic disorder caused by mutations in the *MECP2* (methyl CpG binding protein 2) gene. RTT is no longer listed under “neurodevelopmental disorders” in the DSM-5. However, it is still widely regarded in the scientific and clinical communities as a neurodevelopmental condition due to its early onset, developmental regression, and profound effects on brain maturation. Moreover, the ICD-11 classifies RTT under

“Conditions with disorders of intellectual development as a relevant clinical feature”(LD90.4).

While RTT shares clinical features and gut microbial traits with ASD, the role of Trp derivatives in the disorder is underexplored [149]. Early findings indicate that postmortem brains of RTT patients have increased KYN in specific areas, including the putamen, caudate nucleus, globus pallidus, raphe, and amygdaloid nucleus. 5-HT and 5-HIAA were decreased compared to healthy subjects [96]. Nonetheless, a study on 34 RTT patients found decreased plasma KYN levels and lower bacterial-derived Trp metabolites, indolepropionate and indolelactate—typically produced by *Clostridium sporogenes* species—compared to age- and gender-matched siblings. However, Trp was not altered in RTT subjects [98]. Peripheral reduction contrasts with the brain-specific increase in KYN, highlighting the need to understand regional Trp metabolism rather than relying on systemic measures.

In *Mecp2* knockout mice, decreased 5-HT and serotonergic receptors (*Htr2a*, *Htr3a*) are linked to motor impairments [97]. In the RTT mouse brain, lower LAT1 transporter expression may limit Trp crossing the BBB, disrupting neural catabolic pathways [150]. Yet, whether this is a downstream effect of MeCP2 dysfunction or an independent co-pathology remains unclear. Moreover, the role of LAT1 in selectively limiting Trp and no other amino acids has not been directly validated in RTT models.

Alterations in Trp metabolism seem to be independent of specific *Mecp2* mutations, with similar outcomes observed in mouse models of MeCP2 duplication syndrome, caused by an extra copy of the gene and characterized by intellectual disability and autistic-like phenotypes [99]. Metabolic profiles of this model showed significant changes in 5-HT levels, with increased serum 5-HT and decreased levels in urine and cecum compared to WT controls. Divergent 5-HT levels may reflect altered transport or compartmentalized metabolism rather than synthesis, highlighting the importance of metabolite dynamics over absolute levels. Finally, microbial profiling revealed an altered gut microbiome, with decreased *Firmicutes* (*Lachnospiraceae* spp., *Roseburia* spp., *Dorea* spp., and *Acetivibrio* spp.) and increased *Bacteroides* (*Bacteroides* spp. and *Parabacteroides* sp.). These shifts in microbial composition led to reduced levels of various indole and indole derivatives in plasma samples, including tryptamine, 4-hydroxyindole, 3-indolepropionic acid, indole-3-lactic acid (ILA), and 5-hydroxyindoleacetic acid [99]. Acting as AhR ligands, diminished levels of these indole derivatives could have consequences for multiple metabolic processes, including intestinal barrier integrity and inflammatory responses. However, in vivo functional studies directly linking decreased microbial indole production to RTT symptoms remain limited.

Despite these promising findings, research investigating the link between RTT and Trp metabolism is limited, warranting further investigation into its role in pathogenesis and comorbidities.

Summary of altered tryptophan (Trp) metabolites is reported in autism spectrum disorder (ASD), attention-deficit/hyperactivity disorder (ADHD), Rett syndrome (RTT), and MeCP2 duplication syndrome. For each metabolite, the direction of change (\uparrow or \downarrow), affected tissue, and evidence from preclinical and/or clinical studies are reported. Pre-clinical evidence refers to findings from animal models or in vitro systems, while clinical evidence includes studies involving biological samples from human patients. References correspond to those cited in the main text. The literature search was conducted on PubMed and Consensus (<https://consensus.app/>) using combinations of the keywords: “tryptophan metabolism,” “kynurenine,” “serotonin,” “indole,” AND each disorder name (e.g., “autism,” “ADHD,” “Rett syndrome,” “MeCP2 duplication”), with no time restriction and including both preclinical and clinical studies.

Potential Therapeutic Strategies Targeting Tryptophan Metabolism in Neurodevelopmental Disorders

Trp metabolism is being investigated as a therapeutic target for cancer, neurodegenerative, and autoimmune diseases [22, 151–153]. Approaches include administering metabolites, using probiotics, and targeting key enzymes in 5-HT synthesis and KP (e.g., IDO1, TDO, and KMO) [154]. While most studies focus on modulating the 5-HT system, a few studies have applied therapeutic strategies targeting other Trp metabolic pathways to neurodevelopmental disorders.

Trp administration is used to modulate the serotonergic system [155], with prenatal Trp deficiency reducing serotonergic neurons by 35% [156] underscoring its importance for fetal neurodevelopment and prevention of neurodevelopmental disorders. While supported by preclinical data, human Trp levels are tightly regulated, and high-dose supplementation carries the risk of shunting toward potentially neurotoxic KP metabolites like QA [24, 157]. Defining an optimal Trp dose remains challenging due to variations in brain 5-HT levels across different regions and developmental stages. Additionally, factors such as limited patient availability in clinical trials, difficulties in matching control groups, and individual differences in diet, metabolism, and genetics further complicate standardizing Trp-based interventions.

Pharmacologically, targeting the 5-HT_{1A} receptor has shown promise in treating various symptoms associated with neurodevelopmental disorders [58], including respiratory deficits, anxiety, and stereotypical movements in

ASD and RTT syndrome [158]. Indeed, treatment with the agonist 8-OH-DPAT improved social behavior and helped extinguish fear memory in an ASD rat model [159]. In line with this, low-dose buspirone, a 5-HT_{1A} receptor agonist, has been proposed as an adjunct therapy for restrictive and repetitive behaviors in young children with ASD, alongside behavioral interventions [160]. Mirtazapine, a noradrenergic and serotonergic antidepressant, improves sleep and mood disturbances in both *Mecp2*^{+/-} mice and RTT patients [161]. Selective serotonin reuptake inhibitors (SSRIs) like fluoxetine are commonly prescribed for ASD, with evidence supporting their benefits for psychiatric symptoms in RTT [162], and motor deficits in *Mecp2* mouse models [163]. Clinical results, particularly in ASD, remain inconsistent, with SSRI efficacy varying with age, symptoms, and individual response. Side effects and the lack of stratified protocols currently limit their clinical application [164, 165].

Another potential therapeutic strategy involves modulating the enzyme tryptophan hydroxylase 2 (TPH2), which catalyzes the first and rate-limiting step in the biosynthesis of serotonin [166]. Recent research has resolved the cryo-EM structure of the TPH2 tetramer and identified ZINC000068568685 as a promising small-molecule activator with high binding affinity, laying the groundwork for novel TPH2-targeted drugs [167].

Metformin, a first-line treatment for type 2 diabetes, exhibits neuroprotective effects and may improve ASD behavioral phenotypes [168, 169]. Metformin has been shown to alter Trp, 5-HT, and 5-HIAA levels in the colon and cerebral cortex of high-fat diet-fed BTBR mice, a model of ASD. This may be due to its modulation of Trp metabolism, potentially restoring the 5-HT pathway, although its broad effects on the microbiome, insulin signaling, and mitochondria, obscure whether Trp modulation drives behavioral improvements [170]. Importantly, recent findings suggest that metformin reprograms Trp metabolism by directly influencing the gut microbiome in mice exposed to chronic stress, ultimately alleviating depressive-like behaviors [171].

Probiotic therapy targeting the gut-brain axis has also emerged as a promising intervention for ASD [172, 173]. Daily administration of *Lactobacillus helveticus* CCFM1076 in the VTA rat model of ASD has been reported to restore neurotransmitter homeostasis by improving the balance of the 5-HT system in the peripheral and CNS and ameliorating autistic-like behaviors [174]. Additionally, the probiotic *Bifidobacterium longum* CCFM077 has been reported to restore kynurenine metabolism in ASD rat models, improving autistic-like behaviors, while also regulating levels of QA, glutamic acid, and GABA, and reducing microglial activity in the cerebellum [175]. The QA level in the brain correlated with behavioral improvements, suggesting its potential as a biomarker for autism treatment. Pharmacological or dietary interventions targeting Trp pathways (e.g.,

enzyme inhibitors and probiotics) show promise in modulating neuroinflammation and neurotransmitter balance. While Trp metabolism represents a favorable therapeutic target for various pathological conditions [176], its application to neurodevelopmental disorders is still emerging and largely based on animal studies.

Limitations of the Current Research Landscape

While growing evidence links Trp metabolism to neurodevelopmental disorders, several methodological limitations remain. Most findings come from animal models, post-mortem tissue, or peripheral biospecimens, which may not reflect human CNS or developmental processes. Human studies are often cross-sectional, limiting causal insights, and rely on in vitro models or non-neural samples like urine or feces, raising concerns about compartment-specific relevance.

Many studies assess only a narrow set of metabolites, overlooking pathway complexity. Transcript-level data on key enzymes (e.g., TPH and IDO) often lack corresponding protein or activity measures. Inconsistencies in platforms, sample handling, and cohort variables (age, sex, diet, and microbiota) hinder comparability.

These limitations highlight the need for future research to adopt standardized, multimodal, and longitudinal designs to elucidate the developmental and clinical implications of disrupted Trp metabolism.

Concluding Remarks and Open Questions

The existing literature underscores the complex interplay between multiple factors—maternal diet, stress exposure, gut health, intestinal microbiota and inflammatory responses—that influence one another and collectively shape Trp metabolism homeostasis. These factors have relevant implications for fetal brain development, with long-lasting effects on the offspring. However, the complexity of Trp metabolism, involving multiple interconnected biochemical pathways that compete for shared substrates and intermediates, continues to hinder a complete understanding of its physiological and pathological mechanisms. Several open questions remain: (i) How does the gut microbiota influence host Trp metabolism, limiting Trp availability for absorption, protein synthesis, serotonin, or KP metabolites? (ii) Beyond maternal stress, can postnatal stress alter peripheral and central Trp metabolic pathways, impacting brain network maturation and plasticity during critical periods of development? (iii) Can perinatal dysbiosis, such as prolonged antibiotic use or infant malnutrition disrupt Trp metabolism,

triggering inflammatory responses and altering brain function? (iv) Does an early-life imbalance in the KP or gut microbiota-derived indole production contribute to the onset of ASD and ADHD, or exacerbate neurological symptoms in neurodevelopmental disorders more broadly? Although still in its early stage, research is revealing therapeutic potential. Substantial efforts are required to identify robust biomarkers and enable patient stratification based on metabolic profiles. Looking ahead, the discovery of specific metabolites or targetable molecules (i.e., enzymes and gut microbes) could pave the way for novel interventions. If applied during crucial developmental windows, perhaps beginning as early as the prenatal period, such strategies may counteract the long-term consequences of Trp metabolism disruption on brain function and neurological outcomes.

Acknowledgements We thank all the members of Tognini's team for comments and feedback on the review. Special thanks to Julia Afsar-Keshmiri for her help in editing the manuscript.

Author Contribution MGG and PT conceptualized the review and co-wrote the manuscript. MGG designed the figure, PT supervised and provided the funding.

Funding Open access funding provided by Scuola Superiore Sant'Anna within the CRUI-CARE Agreement. This work was supported by NextGenerationEU Italian Ministry of University and Research PRIN-PNRR2022 CARE P2022CXN7X CUP I53D23006920001, PRIN2022 BAGELS CUP J53C24003080001, Jerome Lejeune Foundation Advanced Grant 2022.

Data Availability No datasets were generated or analysed during the current study.

Declarations

Competing Interests The authors declare no competing interests.

Open Access This article is licensed under a Creative Commons Attribution 4.0 International License, which permits use, sharing, adaptation, distribution and reproduction in any medium or format, as long as you give appropriate credit to the original author(s) and the source, provide a link to the Creative Commons licence, and indicate if changes were made. The images or other third party material in this article are included in the article's Creative Commons licence, unless indicated otherwise in a credit line to the material. If material is not included in the article's Creative Commons licence and your intended use is not permitted by statutory regulation or exceeds the permitted use, you will need to obtain permission directly from the copyright holder. To view a copy of this licence, visit <http://creativecommons.org/licenses/by/4.0/>.

References

- Peters JC (1991) Tryptophan nutrition and metabolism: an overview. *Adv Exp Med Biol* 294:345–358
- Tennoune N, Andriamihaja M, Blachier F (2022) Production of indole and indole-related compounds by the intestinal microbiota and consequences for the host: the good, the bad, and the ugly. *Microorganisms* 10:930
- Jiang H, Chen C, Gao J (2022) Extensive summary of the important roles of indole propionic acid, a gut microbial metabolite in host health and disease. *Nutrients* 15:151
- Pappolla MA, Perry G, Fang X et al (2021) Indoles as essential mediators in the gut-brain axis. Their role in Alzheimer's disease. *Neurobiol Dis* 156:105403
- Alexeev EE, Lanis JM, Kao DJ et al (2018) Microbiota-derived indole metabolites promote human and murine intestinal homeostasis through regulation of interleukin-10 receptor. *Am J Pathol* 188:1183–1194
- Berger M, Gray JA, Roth BL (2009) The expanded biology of serotonin. *Annu Rev Med* 60:355–366
- Guzel T, Mirowska-Guzel D (2022) The role of serotonin neurotransmission in gastrointestinal tract and pharmacotherapy. *Molecules* 27:1680
- Klaessens S, Stroobant V, De Plaen E, Van den Eynde BJ (2022) Systemic tryptophan homeostasis. *Front Mol Biosci* 9:897929
- Gevi F, Zolla L, Gabriele S, Persico AM (2016) Urinary metabolomics of young Italian autistic children supports abnormal tryptophan and purine metabolism. *Mol Autism* 7:47
- Huang X, Ding W, Wu F et al (2020) Increased plasma kynurenic acid levels are associated with impaired attention/vigilance and social cognition in patients with schizophrenia. *Neuropsychiatr Dis Treat* 16:263–271
- Erhardt S, Schwieler L, Imbeault S, Engberg G (2017) The kynurenine pathway in schizophrenia and bipolar disorder. *Neuropharmacology* 112:297–306
- Gulaj E, Pawlak K, Bien B, Pawlak D (2010) Kynurenine and its metabolites in Alzheimer's disease patients. *Adv Med Sci* 55:204–211
- Liang Y, Xie S, He Y et al (2022) Kynurenine pathway metabolites as biomarkers in Alzheimer's disease. *Dis Markers* 2022:9484217
- Lajkó N, Kata D, Szabó M et al (2020) Sensitivity of rodent microglia to kynurenines in models of epilepsy and inflammation *in vivo* and *in vitro*: microglia activation is inhibited by kynurenic acid and the synthetic analogue SZR104. *Int J Mol Sci* 21:9333
- Yan J, Kothur K, Innes EA et al (2022) Decreased cerebrospinal fluid kynurenic acid in epileptic spasms: a biomarker of response to corticosteroids. *EBioMedicine* 84:104280
- Fernandes BS, Inam ME, Enduru N et al (2023) The kynurenine pathway in Alzheimer's disease: a meta-analysis of central and peripheral levels. *Rev Bras Psiquiatr* 45:286–297
- Hargreaves KM, Pardridge WM (1988) Neutral amino acid transport at the human blood-brain barrier. *J Biol Chem* 263:19392–19397
- Bender DA (1983) Biochemistry of tryptophan in health and disease. *Mol Aspects Med* 6:101–197
- Fuchs D, Schroecksadel K, Neurauter G, Bellmann-Weiler R, Ledochowski M, Weiss G (2010) Quality of Life and Tryptophan Degradation. In: Preedy VR, Watson RR (eds) *Handbook of Disease Burdens and Quality of Life Measures*. Springer, New York, NY. https://doi.org/10.1007/978-0-387-78665-0_119
- Moffett JR, Nambodiri MA (2003) Tryptophan and the immune response. *Immunol Cell Biol* 81:247–265
- Jenkins TA, Nguyen JCD, Polglaze KE, Bertrand PP (2016) Influence of tryptophan and serotonin on mood and cognition with a possible role of the gut-brain axis. *Nutrients* 8:56
- Platten M, Nollen EAA, Röhrig UF et al (2019) Tryptophan metabolism as a common therapeutic target in cancer, neurodegeneration and beyond. *Nat Rev Drug Discov* 18:379–401
- Yamamoto T, Azechi H, Board M (2012) Essential role of excessive tryptophan and its neurometabolites in fatigue. *Can J Neurol Sci* 39:40–47

24. Fernstrom JD (2012) Effects and side effects associated with the non-nutritional use of tryptophan by humans. *J Nutr* 142:2236S–2244S
25. Volpi-Abadie J, Kaye AM, Kaye AD (2013) Serotonin syndrome. *Ochsner J* 13:533–540
26. Sun P, Wang M, Chai X et al (2025) Disruption of tryptophan metabolism by high-fat diet-triggered maternal immune activation promotes social behavioral deficits in male mice. *Nat Commun* 16:2105
27. Chajadine M, Laurans L, Radecke T et al (2024) Harnessing intestinal tryptophan catabolism to relieve atherosclerosis in mice. *Nat Commun* 15:6390
28. Sun P, Wang M, Liu Y-X et al (2023) High-fat diet-disturbed gut microbiota-colonocyte interactions contribute to dysregulating peripheral tryptophan-kynurenine metabolism. *Microbiome* 11:154
29. Qi Q, Li J, Yu B et al (2022) Host and gut microbial tryptophan metabolism and type 2 diabetes: an integrative analysis of host genetics, diet, gut microbiome and circulating metabolites in cohort studies. *Gut* 71:1095–1105
30. de Mello VD, Paananen J, Lindström J et al (2017) Indolepropionic acid and novel lipid metabolites are associated with a lower risk of type 2 diabetes in the Finnish Diabetes Prevention Study. *Sci Rep* 7:46337
31. Yang H-L, Feng P, Xu Y et al (2021) The role of dietary fiber supplementation in regulating uremic toxins in patients with chronic kidney disease: a meta-analysis of randomized controlled trials. *J Ren Nutr* 31:438–447
32. Sinha AK, Laursen MF, Brinck JE et al (2024) Dietary fibre directs microbial tryptophan metabolism via metabolic interactions in the gut microbiota. *Nat Microbiol* 9:1964–1978
33. Roager HM, Licht TR (2018) Microbial tryptophan catabolites in health and disease. *Nat Commun* 9:1–10
34. Taleb S (2019) Tryptophan dietary impacts gut barrier and metabolic diseases. *Front Immunol* 10:2113
35. Notarangelo FM, Pocivavsek A (2017) Elevated kynurenine pathway metabolism during neurodevelopment: implications for brain and behavior. *Neuropharmacology* 112:275–285
36. Guidetti P, Amori L, Sapko MT et al (2007) Mitochondrial aspartate aminotransferase: a third kynurenate-producing enzyme in the mammalian brain. *J Neurochem* 102:103–111
37. Stone TW, Darlington LG (2013) The kynurenine pathway as a therapeutic target in cognitive and neurodegenerative disorders: kynurenines and CNS disorders. *Br J Pharmacol* 169:1211–1227
38. Guillemin GJ, Smith DG, Smythe GA et al (2003) Expression of the kynurenine pathway enzymes in human microglia and macrophages. *Adv Exp Med Biol* 527:105–112
39. Perkins MN, Stone TW (1982) An iontophoretic investigation of the actions of convulsant kynurenines and their interaction with the endogenous excitant quinolinic acid. *Brain Res* 247:184–187
40. Esser C (2016) *Suppression and regulation of immune responses: methods and protocols*. Springer, New York, NY
41. Gutiérrez-Vázquez C, Quintana FJ (2018) Regulation of the immune response by the aryl hydrocarbon receptor. *Immunity* 48:19–33
42. Kawajiri K, Fujii-Kuriyama Y (2017) The aryl hydrocarbon receptor: a multifunctional chemical sensor for host defense and homeostatic maintenance. *Exp Anim* 66:75–89
43. Hubbard TD, Murray IA, Bisson WH et al (2015) Adaptation of the human aryl hydrocarbon receptor to sense microbiota-derived indoles. *Sci Rep* 5:12689
44. Barroso A, Mahler JV, Fonseca-Castro PH, Quintana FJ (2021) The aryl hydrocarbon receptor and the gut-brain axis. *Cell Mol Immunol* 18:259–268
45. Coretti L, Buommino E, Lembo F (2024) The aryl hydrocarbon receptor pathway: a linking bridge between the gut microbiome and neurodegenerative diseases. *Front Cell Neurosci* 18:1433747
46. Reyes Ocampo J, Lugo Huitrón R, González-Esquivel D et al (2014) Kynurenines with neuroactive and redox properties: relevance to aging and brain diseases. *Oxid Med Cell Longev* 2014:646909
47. Ma N, He T, Johnston LJ, Ma X (2020) Host-microbiome interactions: the aryl hydrocarbon receptor as a critical node in tryptophan metabolites to brain signaling. *Gut Microbes* 11:1203–1219
48. Yang Y, Wang N, Xu L et al (2023) Aryl hydrocarbon receptor dependent anti-inflammation and neuroprotective effects of tryptophan metabolites on retinal ischemia/reperfusion injury. *Cell Death Dis* 14:92
49. Kimura E, Ding Y, Tohyama C (2016) AhR signaling activation disrupts migration and dendritic growth of olfactory interneurons in the developing mouse. *Sci Rep* 6:26386
50. Martin NR, Patel R, Kossack ME et al (2022) Proper modulation of AHR signaling is necessary for establishing neural connectivity and oligodendrocyte precursor cell development in the embryonic zebrafish brain. *Front Mol Neurosci* 15:1032302
51. Joisten N, Ruas JL, Braidy N et al (2021) The kynurenine pathway in chronic diseases: a compensatory mechanism or a driving force? *Trends Mol Med* 27:946–954
52. Huang X, Feng Z, Cheng H-W (2022) Perspective: gestational tryptophan fluctuation altering neuroembryogenesis and psychosocial development. *Cells* 11:1270
53. Badawy AA-B (2015) Tryptophan metabolism, disposition and utilization in pregnancy. *Biosci Rep* 35:e00261
54. Karahoda R, Abad C, Horackova H et al (2020) Dynamics of tryptophan metabolic pathways in human placenta and placental-derived cells: effect of gestation age and trophoblast differentiation. *Front Cell Dev Biol* 8:574034
55. Richard DM, Dawes MA, Mathias CW et al (2009) L-tryptophan: basic metabolic functions, behavioral research and therapeutic indications. *Int J Tryptophan Res* 2:45–60
56. Russo S, Kema IP, Bosker F et al (2009) Tryptophan as an evolutionarily conserved signal to brain serotonin: molecular evidence and psychiatric implications. *World J Biol Psychiatry* 10:258–268
57. Li D, Yu S, Long Y et al (2022) Tryptophan metabolism: mechanism-oriented therapy for neurological and psychiatric disorders. *Front Immunol* 13:985378
58. Celada P, Bortolozzi A, Artigas F (2013) Serotonin 5-HT_{1A} receptors as targets for agents to treat psychiatric disorders: rationale and current status of research. *CNS Drugs* 27:703–716
59. Murakami Y, Imamura Y, Kasahara Y et al (2023) Maternal inflammation with elevated kynurenine metabolites is related to the risk of abnormal brain development and behavioral changes in autism spectrum disorder. *Cells* 12:1087
60. Cohen Kadosh K, Muhandi L, Parikh P et al (2021) Nutritional support of neurodevelopment and cognitive function in infants and young children—an update and novel insights. *Nutrients* 13:199
61. Sedlmayr P, Blaschitz A, Stocker R (2014) The role of placental tryptophan catabolism. *Front Immunol* 5:230
62. Laurent L, Deroy K, St-Pierre J et al (2017) Human placenta expresses both peripheral and neuronal isoform of tryptophan hydroxylase. *Biochimie* 140:159–165
63. Murthi P, Wallace EM, Walker DW (2017) Altered placental tryptophan metabolic pathway in human fetal growth restriction. *Placenta* 52:62–70
64. Rosenfeld CS (2021) The placenta-brain-axis. *J Neurosci Res* 99:271–283

65. van Zundert SKM, van Egmond NCM, van Rossem L et al (2024) First trimester maternal tryptophan metabolism and embryonic and fetal growth: the Rotterdam Periconceptional Cohort (Predict Study). *Hum Reprod* 39:912–922
66. Tsuji A, Nakata C, Sano M et al (2013) L-tryptophan metabolism in pregnant mice fed a high L-tryptophan diet and the effect on maternal, placental, and fetal growth. *Int J Tryptophan Res* 6:21–33
67. Xu K, Liu H, Bai M et al (2017) Redox properties of tryptophan metabolism and the concept of tryptophan use in pregnancy. *Int J Mol Sci*. <https://doi.org/10.3390/ijms18071595>
68. Okuda S, Nishiyama N, Saito H, Katsuki H (1998) 3-hydroxykynurenine, an endogenous oxidative stress generator, causes neuronal cell death with apoptotic features and region selectivity. *J Neurochem* 70:299–307
69. Moura CA, Cagni FC, Costa LRF et al (2022) Maternal stress during pregnancy in mice induces sex-dependent behavioral alterations in offspring along with impaired serotonin and kynurenine pathways of tryptophan metabolism. *Dev Neurosci* 44:603–614
70. Marszalek-Grabska M, Gawel K, Kosheva N et al (2023) Developmental exposure to kynurenine affects zebrafish and rat behavior. *Cells*. <https://doi.org/10.3390/cells12182224>
71. Meyer U, Feldon J (2012) To poly(I:C) or not to poly(I:C): advancing preclinical schizophrenia research through the use of prenatal immune activation models. *Neuropharmacology* 62:1308–1321
72. Meyer U, Feldon J, Schedlowski M, Yee BK (2005) Towards an immuno-precipitated neurodevelopmental animal model of schizophrenia. *Neurosci Biobehav Rev* 29:913–947
73. Goeden N, Velasquez J, Arnold KA et al (2016) Maternal inflammation disrupts fetal neurodevelopment via increased placental output of serotonin to the fetal brain. *J Neurosci* 36:6041–6049
74. Abad C, Karahoda R, Orbisova A et al (2024) Pathological shifts in tryptophan metabolism in human term placenta exposed to LPS or poly I:C. *Biol Reprod* 110:722–738
75. Karahoda R, Robles M, Marushka J et al (2021) Prenatal inflammation as a link between placental expression signature of tryptophan metabolism and preterm birth. *Hum Mol Genet* 30:2053–2067
76. Frerichs NM, de Meij TGJ, Niemarkt HJ (2024) Microbiome and its impact on fetal and neonatal brain development: current opinion in pediatrics. *Curr Opin Clin Nutr Metab Care* 27:297–303
77. Bresesti I, Salvatore S, Valetti G et al (2022) The microbiota-gut axis in premature infants: physio-pathological implications. *Cells* 11:379
78. Kim E, Paik D, Ramirez RN et al (2022) Maternal gut bacteria drive intestinal inflammation in offspring with neurodevelopmental disorders by altering the chromatin landscape of CD4+ T cells. *Immunity* 55:145–158.e7
79. Galley JD, Chen HJ, Antonson AM, Gur TL (2021) Prenatal stress-induced disruptions in microbial and host tryptophan metabolism and transport. *Behav Brain Res* 414:113471
80. Galley JD, King MK, Rajasekera TA et al (2024) Gestational administration of *Bifidobacterium dentium* results in intergenerational modulation of inflammatory, metabolic, and social behavior. *Brain Behav Immun* 122:44–57
81. Sugino KY, Janssen RC, McMahan RH et al (2024) Vertical transfer of maternal gut microbes to offspring of Western diet-fed dams drives reduced levels of tryptophan metabolites and postnatal innate immune response. *Nutrients* 16:1808
82. Asp L, Holtze M, Powell SB et al (2010) Neonatal infection with neurotropic influenza A virus induces the kynurenine pathway in early life and disrupts sensorimotor gating in adult Tap1^{-/-} mice. *Int J Neuropsychopharmacol* 13:475–485
83. Ramirez Ortega D, Ovalle Rodríguez P, Pineda B et al (2020) Kynurenine pathway as a new target of cognitive impairment induced by lead toxicity during the lactation. *Sci Rep* 10:3184
84. Réus GZ, Silva RH, de Moura AB et al (2019) Early maternal deprivation induces microglial activation, alters glial fibrillary acidic protein immunoreactivity and indoleamine 2,3-dioxygenase during the development of offspring rats. *Mol Neurobiol* 56:1096–1108
85. Pocivavsek A, Wu H-Q, Elmer GI et al (2012) Pre- and postnatal exposure to kynurenine causes cognitive deficits in adulthood. *Eur J Neurosci* 35:1605–1612
86. Fung TC, Olson CA, Hsiao EY (2017) Interactions between the microbiota, immune and nervous systems in health and disease. *Nat Neurosci* 20:145–155
87. Lapin IP (1978) Stimulant and convulsive effects of kynurenines injected into brain ventricles in mice. *J Neural Transm* 42:37–43
88. Schwarcz R, Brush GS, Foster AC, French ED (1984) Seizure activity and lesions after intrahippocampal quinolinic acid injection. *Exp Neurol* 84:1–17
89. Nakano K, Takahashi S, Mizobuchi M et al (1993) High levels of quinolinic acid in brain of epilepsy-prone E1 mice. *Brain Res* 619:195–198
90. Guo J, Williams DJ, Puhl HL 3rd, Ikeda SR (2008) Inhibition of N-type calcium channels by activation of GPR35, an orphan receptor, heterologously expressed in rat sympathetic neurons. *J Pharmacol Exp Ther* 324:342–351
91. Mu C, Choudhary A, Mayengbam S et al (2022) Seizure modulation by the gut microbiota and tryptophan-kynurenine metabolism in an animal model of infantile spasms. *EBioMedicine* 76:103833
92. Hanin A, Chollet C, Demeret S et al (2024) Metabolomic changes in adults with status epilepticus: a human case-control study. *Epilepsia* 65:929–943
93. Chen Y, Guillemin GJ (2009) Kynurenine pathway metabolites in humans: disease and healthy states. *Int J Tryptophan Res* 2:1–19
94. Issac A, Halemani K, Shetty A et al (2025) The global prevalence of autism spectrum disorder in children: a systematic review and meta-analysis. *Osong Public Health Res Perspect* 16:3–27
95. Dobrosavljevic M, Solares C, Cortese S et al (2020) Prevalence of attention-deficit/hyperactivity disorder in older adults: a systematic review and meta-analysis. *Neurosci Biobehav Rev* 118:282–289
96. Rong Y, Yang C-J, Jin Y, Wang Y (2021) Prevalence of attention-deficit/hyperactivity disorder in individuals with autism spectrum disorder: a meta-analysis. *Res Autism Spectr Disord* 83:101759
97. Lewis N, Villani A, Lagopoulos J (2025) Gut dysbiosis as a driver of neuroinflammation in attention-deficit/hyperactivity disorder: a review of current evidence. *Neuroscience* 569:298–321
98. Doenyas C (2018) Gut microbiota, inflammation, and probiotics on neural development in autism spectrum disorder. *Neuroscience* 374:271–286
99. Onore C, Careaga M, Ashwood P (2012) The role of immune dysfunction in the pathophysiology of autism. *Brain Behav Immun* 26:383–392
100. Han VX, Patel S, Jones HF, Dale RC (2021) Maternal immune activation and neuroinflammation in human neurodevelopmental disorders. *Nat Rev Neurol* 17:564–579
101. Hours C, Recasens C, Baleyte J-M (2022) ASD and ADHD comorbidity: what are we talking about? *Front Psychiatry* 13:837424
102. Mostafa GA, Al-Ayadhi LY (2011) A lack of association between hyperserotonemia and the increased frequency of serum anti-myeelin basic protein auto-antibodies in autistic children. *J Neuroinflammation* 8:71

103. Park JH (2022) Potential inflammatory biomarker in patients with attention deficit hyperactivity disorder. *Int J Mol Sci* 23:13054
104. Almulla AF, Thipakorn Y, Tunvirachaisakul C, Maes M (2023) The tryptophan catabolite or kynurenine pathway in autism spectrum disorder; a systematic review and meta-analysis. *Autism Res* 16:2302–2315
105. Neul JL, Skinner SA, Annese F et al (2020) Metabolic signatures differentiate Rett syndrome from unaffected siblings. *Front Integr Neurosci* 14:7
106. Abdulmir HA, Abdul-Rasheed OF, Abdulghani EA (2018) Serotonin and serotonin transporter levels in autistic children. *Saudi Med J* 39:487–494
107. Chakraborti B, Verma D, Guhathakurta S et al (2020) Gender-specific effect of 5-HT and 5-HIAA on threshold level of behavioral symptoms and sex-bias in prevalence of autism spectrum disorder. *Front Neurosci* 13:1375. <https://doi.org/10.3389/fnins.2019.01375>
108. Manzella CR, Jayawardena D, Pagani W et al (2020) Serum serotonin differentiates between disease activity states in Crohn's patients. *Inflamm Bowel Dis* 26:1607–1618
109. Linden DR, Chen J-X, Gershon MD et al (2003) Serotonin availability is increased in mucosa of guinea pigs with TNBS-induced colitis. *Am J Physiol Gastrointest Liver Physiol* 285:G207–G216
110. Grondin JA, Khan WI (2024) Emerging roles of gut serotonin in regulation of immune response, microbiota composition and intestinal inflammation. *J Can Assoc Gastroenterol* 7:88–96
111. Li T, Fu B, Zhang X et al (2021) Overproduction of gastrointestinal 5-HT promotes colitis-associated colorectal cancer progression via enhancing NLRP3 inflammasome activation. *Cancer Immunol Res* 9:1008–1023
112. Banks WA, Kastin AJ, Broadwell RD (1995) Passage of cytokines across the blood-brain barrier. *NeuroImmunoModulation* 2:241–248
113. Blatteis CM (1990) Neuromodulative actions of cytokines. *Yale J Biol Med* 63:133–146
114. Hickey WF, Hsu BL, Kimura H (1991) T-lymphocyte entry into the central nervous system. *J Neurosci Res* 28:254–260
115. Banks WA, Kastin AJ, Gutierrez EG (1993) Interleukin-1 alpha in blood has direct access to cortical brain cells. *Neurosci Lett* 163:41–44
116. Izumi M, Nakanishi Y, Kang S, Kumanogoh A (2024) Peripheral and central regulation of neuro-immune crosstalk. *Inflamm Regen* 44:41
117. Cao H, Tang J, Liu Q et al (2022) Autism-like behaviors regulated by the serotonin receptor 5-HT2B in the dorsal fan-shaped body neurons of *Drosophila melanogaster*. *Eur J Med Res* 27:203
118. Kondaurova EM, Belokopytova II, Kulikova EA et al (2023) On the role of serotonin 5-HT1A receptor in autistic-like behavior: cross talk of 5-HT and BDNF systems. *Behav Brain Res* 438:114168
119. Higazi AM, Kamel HM, Abdel-Naeem EA et al (2021) Expression analysis of selected genes involved in tryptophan metabolic pathways in Egyptian children with autism spectrum disorder and learning disabilities. *Sci Rep* 11:6931
120. Boccutto L, Chen C-F, Pittman AR et al (2013) Decreased tryptophan metabolism in patients with autism spectrum disorders. *Mol Autism* 4:16
121. Zikopoulos B, Barbas H (2013) Altered neural connectivity in excitatory and inhibitory cortical circuits in autism. *Front Hum Neurosci* 7:609
122. Liu X, Bautista J, Liu E, Zikopoulos B (2020) Imbalance of laminar-specific excitatory and inhibitory circuits of the orbitofrontal cortex in autism. *Mol Autism* 11:83
123. Mehra A, Arora G, Sahni G et al (2023) Gut microbiota and autism spectrum disorder: from pathogenesis to potential therapeutic perspectives. *J Tradit Complement Med* 13:135–149
124. Yano JM, Yu K, Donaldson GP et al (2015) Indigenous bacteria from the gut microbiota regulate host serotonin biosynthesis. *Cell* 161:264–276
125. Sanidad KZ, Rager SL, Carrow HC et al (2024) Gut bacteria-derived serotonin promotes immune tolerance in early life. *Sci Immunol* 9(93):eadj4775
126. Golubeva AV, Joyce SA, Moloney G et al (2017) Microbiota-related changes in bile acid & tryptophan metabolism are associated with gastrointestinal dysfunction in a mouse model of autism. *EBioMedicine* 24:166–178
127. Hsiao EY, McBride SW, Hsien S et al (2013) Microbiota modulate behavioral and physiological abnormalities associated with neurodevelopmental disorders. *Cell* 155:1451–1463
128. Flemming A (2024) Bacterial serotonin induces Treg cells in neonates. *Nat Rev Immunol* 24:306
129. Srikantha P, Mohajeri MH (2019) The possible role of the microbiota-gut-brain-axis in autism spectrum disorder. *Int J Mol Sci* 20:2115
130. American Psychiatric Association (2000) Diagnostic and statistical manual of mental disorders: Text revision. 4th edn. Washington DC. <https://doi.org/10.1176/appi.books.9780890423349Tryptophan>
131. Dunn GA, Nigg JT, Sullivan EL (2019) Neuroinflammation as a risk factor for attention deficit hyperactivity disorder. *Pharmacol Biochem Behav* 182:22–34
132. Du J, Fang L, Dong K, Zhou Z (2025) Exploring the complex relationship between attention deficit hyperactivity disorder and the immune system: a bidirectional Mendelian randomization analysis. *J Affect Disord* 369:854–860
133. Gainetdinov RR, Wetsel WC, Jones SR et al (1999) Role of serotonin in the paradoxical calming effect of psychostimulants on hyperactivity. *Science* 283:397–401
134. Uzbekov M (2018) Monoamines and kynurenine involvement in pathogenetic mechanisms of attention deficit hyperactivity disorder or hyperkinetic syndrome. *Glob J Intellect Dev Disabil* 5. <https://doi.org/10.19080/gjidd.2018.05.555667>
135. Cavaleri D, Crocamo C, Morello P et al (2024) The kynurenine pathway in attention-deficit/hyperactivity disorder: a systematic review and meta-analysis of blood concentrations of tryptophan and its catabolites. *J Clin Med*. <https://doi.org/10.3390/jcm13020583>
136. Sağlam E, Bilgiç A, Abuşoğlu S et al (2021) The role of tryptophan metabolic pathway in children with attention deficit hyperactivity disorder with and without comorbid oppositional defiant disorder and conduct disorder. *Psychiatry Res* 298:113770
137. Murakami Y, Imamura Y, Saito K et al (2019) Altered kynurenine pathway metabolites in a mouse model of human attention-deficit hyperactivity/autism spectrum disorders: a potential new biological diagnostic marker. *Sci Rep* 9:13182
138. Oades RD (2011) An exploration of the associations of pregnancy and perinatal features with cytokines and tryptophan/kynurenine metabolism in children with attention-deficit hyperactivity disorder (ADHD). *Atten Defic Hyperact Disord* 3:301–318
139. Aarsland TIM, Landaas ET, Hegvik T-A et al (2015) Serum concentrations of kynurenines in adult patients with attention-deficit hyperactivity disorder (ADHD): a case-control study. *Behav Brain Funct* 11:36
140. Quintero J, Gutiérrez-Casares JR, Álamo C (2022) Molecular characterisation of the mechanism of action of stimulant drugs lisdexamfetamine and methylphenidate on ADHD neurobiology: a review. *Neurol Ther* 11:1489–1517

141. Qu S, Zhou X, Wang Z et al (2024) The effects of methylphenidate and atomoxetine on *Drosophila* brain at single-cell resolution and potential drug repurposing for ADHD treatment. *Mol Psychiatry* 29:165–185
142. Molina-Carballo A, Cubero-Millán I, Fernández-López L et al (2021) Methylphenidate ameliorates the homeostatic balance between levels of kynurenines in ADHD children. *Psychiatry Res* 303:114060
143. Fernández-López L, Molina-Carballo A, Cubero-Millán I et al (2020) Indole tryptophan metabolism and cytokine S100B in children with attention-deficit/hyperactivity disorder: daily fluctuations, responses to methylphenidate, and interrelationship with depressive symptomatology. *J Child Adolesc Psychopharmacol* 30:177–188
144. Boonchooduang N, Louthrenoo O, Likhitweerawong N et al (2025) Impact of psychostimulants on microbiota and short-chain fatty acids alterations in children with attention-deficit/hyperactivity disorder. *Sci Rep* 15:3034
145. Aresti-Sanz J, Schwalbe M, Pereira RR et al (2021) Stability of methylphenidate under various pH conditions in the presence or absence of gut microbiota. *Pharmaceuticals (Basel)* 14:733
146. Borghi E, Vignoli A (2019) Rett syndrome and other neurodevelopmental disorders share common changes in gut microbial community: a descriptive review. *Int J Mol Sci* 20:4160
147. Riederer P, Weiser M, Wichart I et al (1986) Preliminary brain autopsy findings in prodromal Rett syndrome. *Am J Med Genet Suppl* 1:305–315
148. Santos M, Summavielle T, Teixeira-Castro A et al (2010) Monoamine deficits in the brain of methyl-CpG binding protein 2 null mice suggest the involvement of the cerebral cortex in early stages of Rett syndrome. *Neuroscience* 170:453–467
149. Illescas S, Diaz-Osorio Y, Serradell A et al (2024) Metabolic characterization of neurogenetic disorders involving glutamatergic neurotransmission. *J Inherit Metab Dis* 47:551–569
150. Wu J, Hu Q, Rao X et al (2024) Gut microbiome and metabolic profiles of mouse model for MeCP2 duplication syndrome. *Brain Res Bull* 206:110862
151. Opitz CA, Somarribas Patterson LF, Mohapatra SR et al (2020) The therapeutic potential of targeting tryptophan catabolism in cancer. *Br J Cancer* 122:30–44
152. Saidi O, Rochette E, Doré É et al (2020) Randomized double-blind controlled trial on the effect of proteins with different tryptophan/large neutral amino acid ratios on sleep in adolescents: the PROTOMORPHEUS study. *Nutrients* 12:1885
153. Peyraud F, Guegan J-P, Bodet D et al (2022) Targeting tryptophan catabolism in cancer immunotherapy era: challenges and perspectives. *Front Immunol* 13:807271
154. Naing A, Eder JP, Piha-Paul SA et al (2020) Preclinical investigations and a first-in-human phase I trial of M4112, the first dual inhibitor of indoleamine 2,3-dioxygenase 1 and tryptophan 2,3-dioxygenase 2, in patients with advanced solid tumors. *J Immunother Cancer* 8:e000870
155. Carneiro IBC, Toscano AE, Lacerda DC et al (2018) L-tryptophan administration and increase in cerebral serotonin levels: systematic review. *Eur J Pharmacol* 836:129–135
156. Flores-Cruz GM, Escobar A (2012) Reduction of serotonergic neurons in the dorsal raphe due to chronic prenatal administration of a tryptophan-free diet. *Int J Dev Neurosci* 30:63–67
157. Fernstrom JD (2016) A perspective on the safety of supplemental tryptophan based on its metabolic fates. *J Nutr* 146:2601S–2608S
158. Abdala AP, Bissonnette JM, Newman-Tancredi A (2014) Pinpointing brainstem mechanisms responsible for autonomic dysfunction in Rett syndrome: therapeutic perspectives for 5-HT1A agonists. *Front Physiol* 5:205
159. Wang C-C, Lin H-C, Chan Y-H et al (2013) 5-HT1A-receptor agonist modified amygdala activity and amygdala-associated social behavior in a valproate-induced rat autism model. *Int J Neuropsychopharmacol* 16:2027–2039
160. Chugani DC, Chugani HT, Wiznitzer M et al (2016) Efficacy of low-dose buspirone for restricted and repetitive behavior in young children with autism spectrum disorder: a randomized trial. *J Pediatr* 170(45–53):e1–4
161. Flores Gutiérrez J, De Felice C, Natali G et al (2020) Protective role of mirtazapine in adult female Mecp2^{+/-} mice and patients with Rett syndrome. *J Neurodev Disord* 12:26
162. Persico AM, Ricciardello A, Cucinotta F (2019) The psychopharmacology of autism spectrum disorder and Rett syndrome. *Handb Clin Neurol* 165:391–414
163. Villani C, Sacchetti G, Carli M, Invernizzi RW (2020) Fluoxetine rescues rotarod motor deficits in Mecp2 heterozygous mouse model of Rett syndrome via brain serotonin. *Neuropharmacology* 176:108221
164. Williams K, Brignell A, Randall M et al (2013) Selective serotonin reuptake inhibitors (SSRIs) for autism spectrum disorders (ASD). *Cochrane Database Syst Rev*. <https://doi.org/10.1002/14651858.CD004677.pub3>
165. Reiersen AM, Handen B (2011) Commentary on “Selective serotonin reuptake inhibitors (SSRIs) for autism spectrum disorders (ASD).” *Evid Based Child Health* 6:1082–1085
166. Kulikova EA, Kulikov AV (2019) Tryptophan hydroxylase 2 as a therapeutic target for psychiatric disorders: focus on animal models. *Expert Opin Ther Targets* 23:655–667
167. Zhu K, Liu C, Gao Y et al (2022) Cryo-EM structure and activator screening of human tryptophan hydroxylase 2. *Front Pharmacol* 13:907437
168. Gantois I, Khoutorsky A, Popic J et al (2017) Metformin ameliorates core deficits in a mouse model of fragile X syndrome. *Nat Med* 23:674–677
169. Wang L, Cai Y, Fan X (2018) Metformin administration during early postnatal life rescues autistic-like behaviors in the BTBR T+ *Ipr3^{tf}/J* mouse model of autism. *Front Behav Neurosci* 12:290
170. Deng W, Li F, Ke H et al (2022) Effect of metformin in autistic BTBR T+ *Ipr3^{tf}/J* mice administered a high-fat diet. *Brain Res Bull* 183:172–183
171. Xie X, Li W, Xiong Z et al (2025) Metformin reprograms tryptophan metabolism via gut microbiome-derived bile acid metabolites to ameliorate depression-like behaviors in mice. *Brain Behav Immun* 123:442–455
172. Patusco R, Ziegler J (2018) Role of probiotics in managing gastrointestinal dysfunction in children with autism spectrum disorder: an update for practitioners. *Adv Nutr* 9:637–650
173. Yang Y, Tian J, Yang B (2018) Targeting gut microbiome: a novel and potential therapy for autism. *Life Sci* 194:111–119
174. Kong Q, Wang B, Tian P et al (2021) Daily intake of *Lactobacillus* alleviates autistic-like behaviors by ameliorating the 5-hydroxytryptamine metabolic disorder in VPA-treated rats during weaning and sexual maturation. *Food Funct* 12:2591–2604
175. Kong Q, Chen Q, Mao X et al (2022) *Bifidobacterium longum* CCFM1077 ameliorated neurotransmitter disorder and neuroinflammation closely linked to regulation in the kynurenine pathway of autistic-like rats. *Nutrients* 14:1615
176. Modoux M, Rolhion N, Mani S, Sokol H (2021) Tryptophan metabolism as a pharmacological target. *Trends Pharmacol Sci* 42:60–73

Publisher's Note Springer Nature remains neutral with regard to jurisdictional claims in published maps and institutional affiliations.